# Contribution of GCF to the Saliva Proteome: Quantitative Proteomic Analysis

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97.659

7.996

893

74

142

11

31

Q73KY2

115/118 Normalized

-4.29

-3.93

-3.38

-3 28

-3.03

-3.02

-2.97

-2.80

-2.75

-2.64

2.76

2.77

2.79

2.83

2.94

3.00

3.23

3.26

0.22

0.28

0 4 1

0 44

0.53

0.53

0.55

0.62

0.64

0.69

29.33

29.55

29.77

30.76

33.11

34.48

40.39

41.30

45.80

60.18

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### Overview

· Saliva is a complex biological fluid containing proteins derived from the salivary glands, gingival crevicular fluid (GCF) and bacteria. It is a rich source of protein biomarkers.

· Here we ask: What proteins in saliva are derived from GCF?

#### Introduction

· Saliva is a complex biological fluid with a large dynamic range. It is a rich source of proteins and collection is non-invasive. However the proportion of GCF derived proteins is currently noorly defined

 Tandem mass spectrometry is one of the most powerful tools for analysing complex protein mixtures. High mass accuracy and fast acquisition speed allow hundreds of peptides to be identified in less than an hour

 We show quantitative online data-dependent liquid chromatography (LC) MS/MS analysis of saliva from healthy dentate and edentulous volunteers performed on a Thermo Fisher Scientific LTQ-Orbitrap Velos mass spectrometer.

· The results show that over 950 proteins were identified with 2 or more peptides. However, none of the peptides were solely observed in the dentate sample.

#### Method

 Saliva was collect from 10 healthy dentate volunteers and 10 healthy edentulous volunteers. • The samples were centrifuged for 10 minutes and the supernatant retained. 10 uL of each sample for dentate and edentulous was combined to give two 100 µL samples.

· The proteins in both samples were reduced and alkylated prior to overnight digestion with Lvs-C and trypsin.

• The samples were labelled with two iTRAQ (ABSciex) labels from an 8-plex (115 and 118) and combined.

· The peptide mixture was fractionated by strong cation exchange (SCX) chromatography. SCX was performed with a Polysulfoethyl A column. Samples were eluted over a 60 minute salt gradient from 0-50% 500 mM KCl (pH 3). 16 fractions were collected.

• The fractions were loaded onto a 75 μm C18 reversed phase analytical column (LC Packings). Peptides were separated over a 30 minute gradient from 3.2 to 44% acetonitrile (0.1% formic acid).

 Samples were infused by use of an Advion Triversa Nanomate nanospray ionization source into a Thermo Fisher Scientific LTQ-Orbitrap Velos hybrid mass spectrometer.

• The mass spectrometer performed an initial high resolution survey scan in the Orbitrap and the three most intense multiply charged ions were selected for collision induced dissociation (CID) tandem mass spectrometry, detected in the ion trap. The same three ions were the fragmented by higher energy collisional dissociation (HCD), detected in the Orbitrap.

· Analysed peptides were placed on an exclusion list for 60 seconds.

• Data were collected with Xcalibur 2.1 (Thermo Fisher Scientific) and analysed using the SEQUEST search algorithm and the IPI human database (v3.75) supplemented with known oral bacteria, concatenated with the reverse sequences.

· CID spectra were used to identify the peptides and HCD spectra were used for quantification.

#### Conclusions

• We have identified over 950 proteins from saliva with two or more peptides including 74 bacterial proteins (derived from P.gingivalis and T.denticola.) at a 1% FDR.

• 97% of the proteins were identified in greater abundance from the healthy dentate saliva. After normalisation 35 proteins were identified with greater than 2 fold (log2) increase

(dentate:edentulous). 29 proteins were identified with greater abundance in the edentulous. None of the proteins identified were only present in the dentate saliva suggesting the GCF

proteins observed in edentulous saliva may be contributed from tissue transudate.

 We have previously identified 31 of these proteins in GCF suggesting they are either tissue transudates or saliva contaminants in GCF.



Protein

Synaptosomal associated protein, 23 kd

Chromosome X open reading frame 3

ATP-binding cassette sub-family F member 2

Glucocorticoid receptor DNA binding factor 1

Titin

MORN repeat containing 1

Cell cycle checkpoint kinase

Interleukin 9 receptor

Zinc finger protein 609

Zinc finger protein 92

Homeobox C12

(115/118).

Amino acid kinase family protein

Organic anion transporter F

Organic anion transporter 2

H2A histone family member V

Iroquois homeobox protein 6

Number of CID events

(dentate/edentulous)

(dentate/edentulous)

Number of peptides identified

Human proteins x2 upregulated

Bacterial proteins x2 upregulated

Number of human proteins identified

Number of bacterial proteins identified

Proteins previously identified from GCF

Table 2: The number of CID events, proteins and peptides identified

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from the LC-MS/MS analysis of dentate and edentulous saliva.

Glutaminyl peptide cyclotransferase

Family with sequence similarity 83, member H IPI:IPI00784320.3

SLC2A4 regulator

Mucin 16

Figure 1: The total ion chromatogram of one of the SCX fractions.



Figure 3: The HCD mass spectrum of the triply charged peptide ion



HCD

Figure 4: The log (2) curve of the ratios of proteins identified in dentate and edentulous saliva.





Figure 6: The extracted protein-protein interaction maps for titin (more abundant in edentulous) a key component of muscle tissue.

Figure 5: The protein-protein interaction map for the identified proteins.

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s<sup>1</sup>v<sup>1</sup>T<sup>1</sup>E<sup>1</sup>Q<sup>2</sup>G<sup>2</sup>A<sup>2</sup>E<sup>2</sup>L<sup>2</sup>S<sup>2</sup>N<sub>E</sub>E<sub>E</sub>R

CID

8.32:1

3 167-4

[iTRAQ-SVTEQGAELSNEER]<sup>3+</sup>. Inset: The m/z region containing the iTRAQ labels.