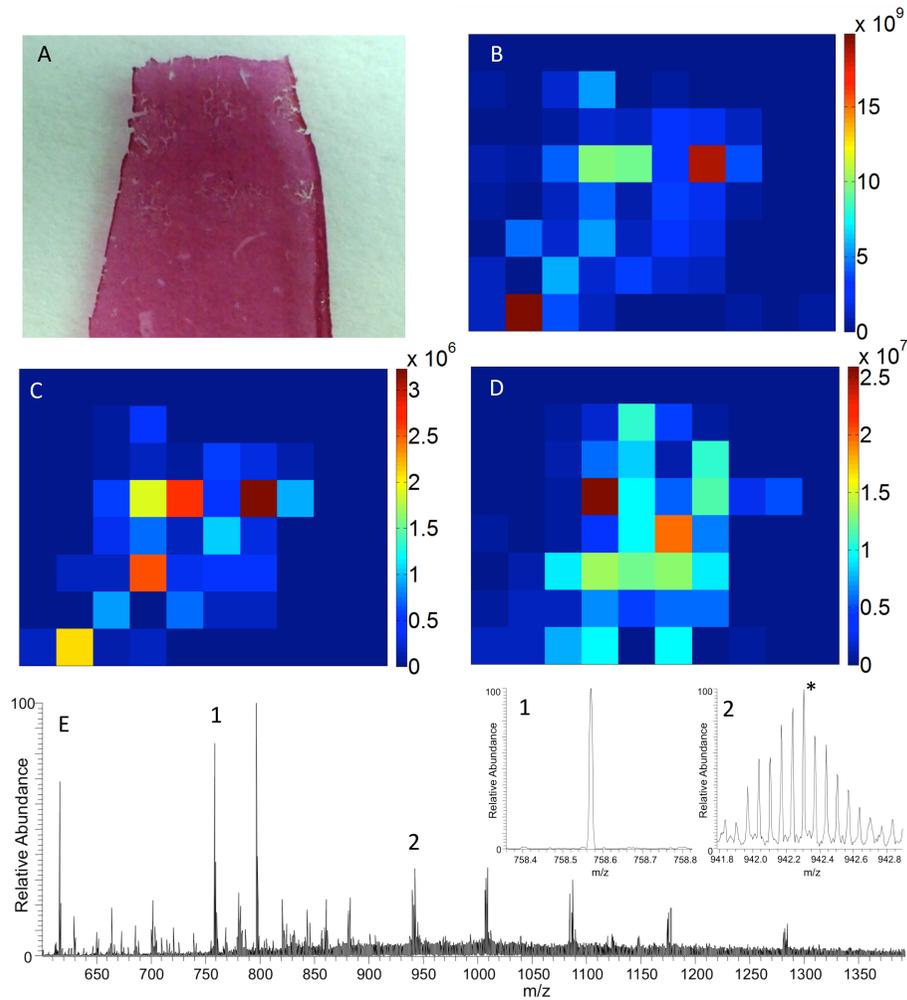


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Introduction:

- Liver Disease is one of the five biggest killers in the UK and is the only one of the five that is increasing year on year [1].
- Liquid extraction surface analysis (LESA) is a new sampling technique in which a liquid micro-junction between a sample and a conductive pipette allows extraction of molecules from a surface, the solution is then injected into the mass spectrometer by chip-based nano-ESI (Advion Nanomate) [2,3].
- LESA has previously been used in analysis of thin layer chromatography plates, dried blood spots, whole animal sections and fresh tissue [3-5]
- In this work we explore the use of LESA coupled to high resolution nano ESI-MS and MS/MS for analysis of lipids and proteins in human liver tissue.
- Opportunities for tissue imaging using in-house software, simultaneous detection of lipids and proteins and the effects of repeated analysis are presented.

Methods:

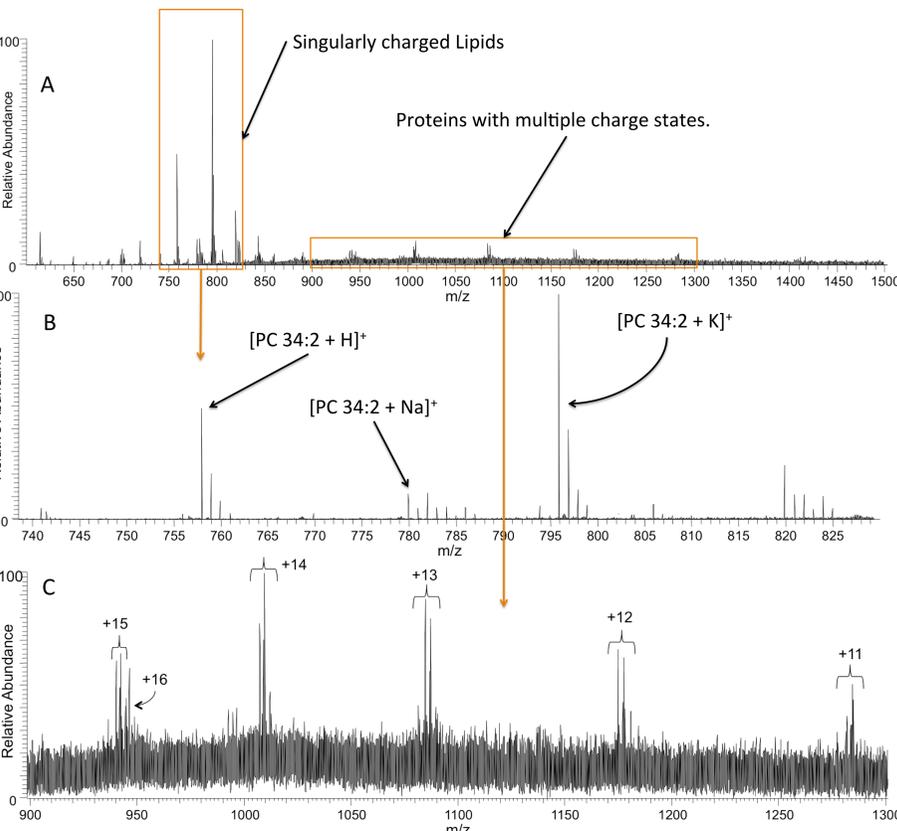
Imaging Lipids and Proteins Liver: LESA sampling - 0.8 μL of 70 % methanol_(aq) with 0.1 % Formic Acid was aspirated from a solvent well of which 0.5 μL was dispensed from a height of 0.2 mm onto tissue sectioned at 10 μm thickness followed by a 10 second delay. 0.5 μL was re-aspirated and electrosprayed into a Velos Orbitrap at a tip voltage of 1.75 kV, gas pressure 0.3 psi and a capillary temperature of 250°C. **MS Method:** All spectra were acquired at a resolution of 100,000 at m/z 400. Each scan comprised 5 co-added microscans and each spectrum was acquired for \sim 15 scans (2 mins). The AGC target was 1×10^6 with a max fill time of 2 s.

Repeat extractions from a single location were performed under similar condition differing only in the solvent volume, 3.5 μL aspirated of which 3 μL was dispensed and reaspirated. And the data was recorded for 3 min at 5 micro scans.

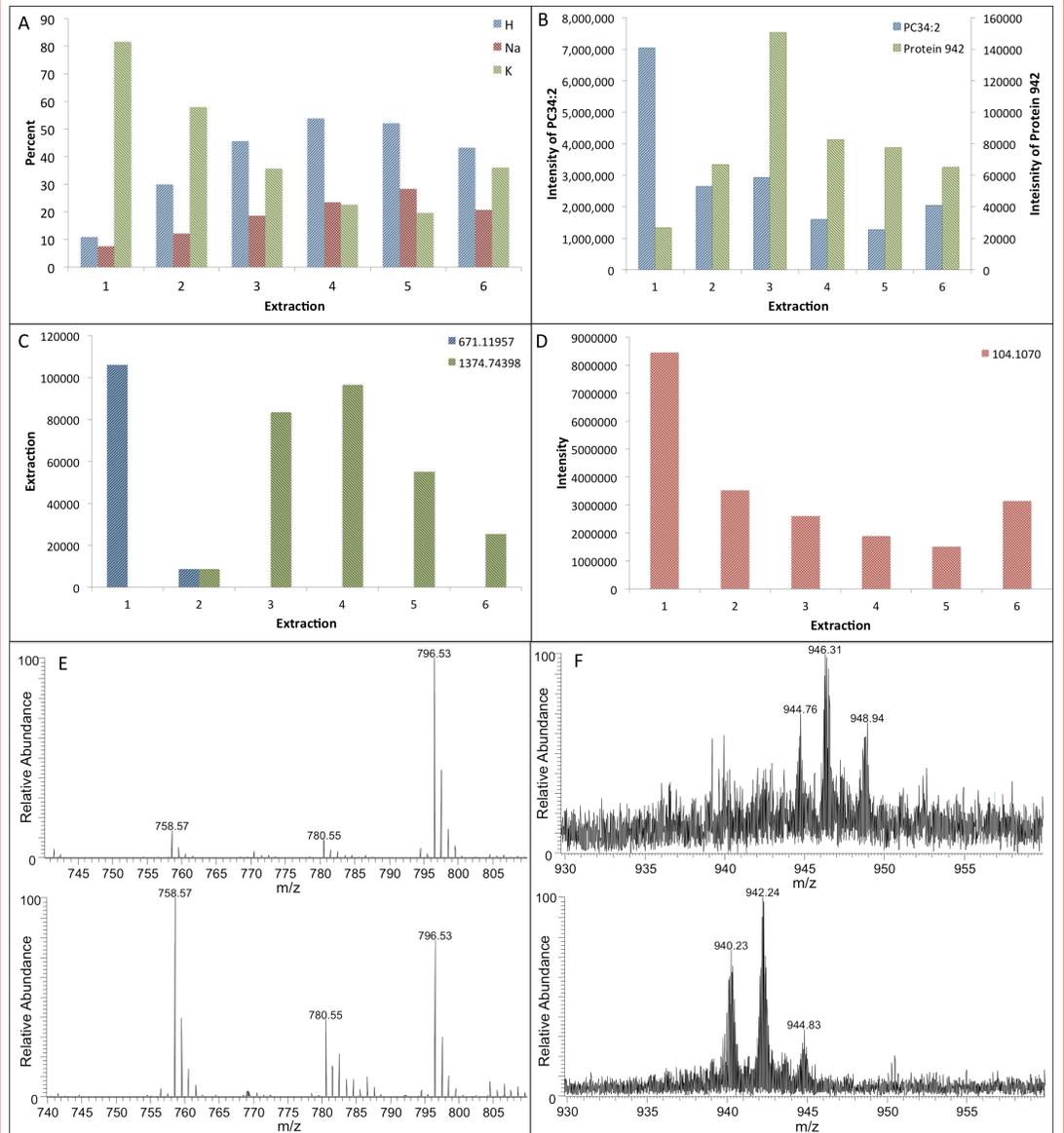
LESA-MS Images from Normal Liver Tissue. A) H&E stain of the tissue after LESA. Localized regions of damage can be seen where the LMJ was maintained. B) Image showing the total ion current from the data acquired. C) m/z 942.3032 (an unidentified protein). D) m/z 758.5676 \pm 0.005 (identified as the protonated adduct of lipid PC 34:2). E) A spectrum from a single pixel on tissue. 1) and 2) indicate the peaks from which the select ion images were taken. In the case of 2) m/z 942.3032, is a single peak (*) from the isotopic distribution from the protein.

Principle Findings:

- Lipids and proteins readily detected in tissue by LESA-MS.
- Direct analysis of tissue via LESA-MS can be used to construct ion images of lipids and proteins in thin tissue sections.
- High mass accuracy and high mass resolving power of the Velos Orbitrap enables identification of some constituents.
- Repeat analysis of the same area enables the detection of additional molecules following the removal of salts and other abundant molecules.



Simultaneous analysis of Lipids and Proteins. A) Typical mass spectrum from liver tissue (m/z 600 – 1500) B) singularity charged lipids detected between m/z 740 and m/z 825. C) Proteins detected as multiple charged species between m/z 900 and m/z 1300.



Repeat Sampling of Tissue via LESA. A) The relative abundance of the lipid PC 34:2 adducts showing the decrease of abundant salts such as potassium and the relative increase of the protonated adduct. B) The absolute intensity of the potassium adduct of PC 34:2 and an unidentified protein m/z 942.3032. C) The 'appearance' and 'disappearance' of two unknown species identified by mass only. D) The depletion of m/z 104.1070 identified as the choline head group from PC lipids. E) Top: first injection, Bottom: third injection. The predominant salt adduct detected changes from $[\text{PC } 34:2 + \text{K}]^+$ to $[\text{PC } 34:2 + \text{H}]^+$. F) Top: first injection, Bottom: third injection. A protein detected at m/z 946.3134 is no longer detected in third injection but a previously undetected protein is observed at m/z 942.2350.

References: [1] British Liver Trust www.britishlivertrust.org.uk/home/about-us/media-centre/facts-about-liver-disease.aspx accessed 10/04/12. [2] Kertesz, V.; Van Berkel, G. J. *Journal of Mass Spectrometry* **2010**, *45*, 252. [3] Edwards, R. L.; Creese, A. J.; Baumert, M.; Griffiths, P.; Bunch, J.; Cooper, H. J. *Analytical Chemistry* **2011**, *83*, 2265. [4] Eikel, D.; Vavrek, M.; Smith, S.; Bason, C.; Yeh, S.; Korfmaier, W. A.; Henion, J. D. *Rapid Communications in Mass Spectrometry* **2011**, *25*, 3587. [5] Walworth, M. J.; Stankovich, J. J.; Van Berkel, G. J.; Schulz, M.; Minarik, S. *Rapid Communications in Mass Spectrometry* **2012**, *26*, 37.

Acknowledgements: To the EPSRC for Funding. To Rebecca Edwards, Cleidiane Zampronion, Andy Creese for useful discussions.